

# EX-PRESS 293™ Cell Lines

For use with EX-PRESS 293™ and EX-PRESS 293™-BL Cells  
Catalog Numbers: MC001, MC001P, MC002, and MC002P  
Version: A1.1 – April 2025  
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## Product Information

### Contents:

Cryovials of frozen cells containing a total of  $1 \times 10^7$  cells in 1 ml of freezing medium (BalanCD HEK293 supplemented with 10% DMSO). Cells are available as single vials, or in packs of 6 vials, see below.

| PRODUCT          | QUANTITY SUPPLIED | CATALOG NUMBER |
|------------------|-------------------|----------------|
| EX-PRESS 293™    | 1 vial            | MC001          |
|                  | 6 vials           | MC001P         |
| EX-PRESS 293™-BL | 1 vial            | MC002          |
|                  | 6 vials           | MC002P         |

**Storage:** The cells are shipped on dry ice. Upon receipt, cryovials should be thawed and used immediately, or transferred to storage in liquid nitrogen. Contact [support@abdesignlabs.com](mailto:support@abdesignlabs.com) if the cells are not frozen upon arrival.

**Quality control:** The cells were certified to meet all specifications, including absence of Mycoplasma species by qPCR screen.

## Product Description

**EX-PRESS 293™** is a HEK293 cell line adapted to be maintained in serum-free media and suspension culture. **EX-PRESS 293™** cells have been optimized for high density, transfection efficiency and recombinant protein production using a protocol designed to reduce transfection requirements without sacrificing recombinant protein production or quality.

**EX-PRESS 293™-BL** is a derivative of the **EX-PRESS 293™** cell line that has been modified for highly efficient in cellulo biotinylation of recombinant proteins that contain an Avi-tag™.

## Required Reagents Not Supplied

- ❖ Sterile 125 ml Optimum Growth® Shake Flasks (Thomson, Cat#931110)
- ❖ BalanCD HEK293 (Fujifilm Irvine Scientific, Cat# 91165)
- ❖ BalanCD HEK293 Feed (Fujifilm Irvine Scientific, Cat# 91166)
- ❖ L-glutamine, 200 mM (any supplier)
- ❖ D-biotin, 50 mM (**EX-PRESS 293™-BL** only, any supplier)
- ❖ Zeocin®, 100 mg/ml solution, (Optional, **EX-PRESS 293™-BL** only, InvivoGen, Cat#ant-zn-05)

## Protocols

### General Cell Culture

#### Handling

All reagents and equipment that come in contact with the cells should be sterile. Work with the cells should be done in a laminar flow hood using proper aseptic technique. Vigorous shaking or pipetting when working with the cells should be avoided to maintain cell health.

#### Cell Thawing

1. Add 30 ml of BalanCD HEK293 supplemented with 2 mM L-glutamine pre-warmed to 37°C in a 125 ml Optimum Growth® shake flask.
2. Transfer a vial of **EX-PRESS 293™** cells from storage in liquid nitrogen to a 37°C water bath and swirl gently for 60 seconds to rapidly thaw the cells. Do not submerge the vial.
3. Before the cells have completely thawed, remove the vial from the water bath and spray the vial thoroughly with 70% ethanol to decontaminate the vial exterior before opening the vial in a laminar flow hood.

**Note:** Incubating the cells too long at 37°C will result in loss of viability.

4. Gently mix the contents of the vial and transfer to the flask prepared previously.
5. Transfer the shake flask into a shaking, humidified incubator set to 37°C, 8% CO<sub>2</sub> and 150 rpm (20 mm orbital diameter).
6. Estimate concentration of cell suspension three to four days post-thaw by hemocytometer or automated cell counter. Viability should be  $\geq 90\%$  with a density  $\geq 1 \times 10^6$  cells/ml. Continue incubation if needed, or proceed to routine subculturing.

## Cell Subculturing

**EX-PRESS 293™** cells have a doubling time of approximately 24 h during log phase growth. Cells should be passaged every 3 to 4 days to maintain density of  $\leq 3 \times 10^6$  viable cells per ml.

1. Estimate concentration of viable cells in suspension by hemocytometer or automated cell counter, and calculate volume required for seeding a new flask at  $3 \times 10^5$  viable cells per ml.

**Note:** We recommend using Optimum Growth® shake flasks for culturing. The unique flask design allows for high volume in a small form factor suitable for our recommended bioprocessing method while maintaining the same shake conditions across flasks of all sizes (125 ml to 7 L).

2. Transfer calculated volume of cell suspension to fresh, pre-warmed, BalanCD HEK293 media supplemented with 2 mM L-glutamine.

**Optional:** **EX-PRESS 293™-BL** cells are resistant to Zeocin® and can be cultured with media supplemented with 100 µg/ml Zeocin® to ensure stable integration of biotin ligase is maintained.

3. Incubate flasks in a shaking, humidified incubator set to 37°C, 8% CO<sub>2</sub> and 150 rpm (20 mm orbital diameter).
4. Repeat steps 1-3 to maintain or expand cells. We recommend discarding cultures after 30 passages.

## Cryopreservation

1. Estimate cell density and culture volume required to preserve the desired number of 1 ml aliquots containing  $1 \times 10^7$  cells per ml.
2. Expand culture up to required volume while maintaining densities  $\leq 3 \times 10^6$  cells/ml and  $\geq 90\%$  viability.
3. Pellet cells by centrifugation at 300 x g for 5 min, aspirate spent medium.
4. Resuspend cell pellet in BalanCD HEK293 media supplemented with 2 mM L-glutamine and 10% DMSO pre-chilled to 4°C at a final concentration of  $1 \times 10^7$  cells/ml.
5. Dispense 1 ml of cell suspension into each cryogenic vial. Place vials in an insulated container and store at -80°C overnight.
6. Transfer vials to liquid nitrogen vapor phase for long-term storage.

## Recombinant Protein Production

### Transfection

**EX-PRESS 293™** cells have been optimized for commercially available cationic-lipid transfection systems but are also compatible with other methods of transfection (e.g., PEI). We recommend researchers follow the manufacturer's protocol for the desired method and reagent.

We recommend testing transfection conditions with a fluorescent reporter plasmid first (i.e., **TGEX™-eGFP-Zeo**, Cat# MX022) to evaluate transfection efficiency and determine optimal transfection conditions.

**TGEX™** series mammalian expression vectors have been optimized for recombinant antibody production in **EX-PRESS 293™** cells, see *Related Products* for available formats and isotypes.

### Fed-batch production

The following protocol is the recommended bioprocessing method using **EX-PRESS 293™** cells.

1. Subculture and expand **EX-PRESS 293™** cells until density reaches  $\geq 3 \times 10^6$  cells/ml in half of the total desired production volume (*Transfection volume*).

**Note:** Do not use cultures with densities  $\geq 3 \times 10^6$  cells/ml for routine maintenance.

2. On the day of transfection, seed **EX-PRESS 293™** cells at  $3 \times 10^6$  cells/ml in the transfection volume in an Optimum Growth® flask with a fill volume suitable for the desired production volume.
3. Transfect culture with 1 µg/ml transfection-grade mammalian expression vector according to the protocol of preferred transfection reagent.

**Note:** For expression of proteins that require co-transfection (e.g., antibody heavy and light chain vectors) we recommend evaluating the ratio of the vectors while maintaining the total DNA concentration at 1 µg/ml.

**Important:** Supplement the media of **EX-PRESS 293™-BL** fed-batch productions with 50 µM D-biotin to facilitate in cellulo biotinylation.

4. Return culture to incubate at 37°C and 8% CO<sub>2</sub> with shaking at 150 rpm (20 mm orbital diameter).
5. If required, enhance cultures with components of the preferred transfection reagent system at times specified in manufacturer's protocol the day after transfection.

6. Two days post-transfection, double the volume of the culture to the final production volume using pre-warmed BalanCD HEK293 supplemented with 2 mM L-glutamine and 50 µM D-biotin (**EX-PRESS 293™-BL** only).

**Note:** Optimum Growth® flasks are recommended as the transfection volume can be doubled to the final production volume in the same flask. Other flasks may be used provided the production volume does not exceed the recommended flask fill volume to maintain optimal gas exchange.

7. Continue to incubate cells, feeding with BalanCD HEK293. Feed as desired according to manufacturer's protocol, for an additional three days before harvesting culture supernatant for downstream purification processes.

**Note:** Optimal harvest time is dependent on the recombinant protein being expressed. We recommend a 6-day fed batch protocol for most secreted proteins.

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## Related Products

| VECTOR   | CATALOG | ISOTYPE/USE          | DESCRIPTION   |
|--|---------|----------------------|---|
| <b>Antibody Heavy Chain Expression Vectors</b>               |         |                      |   |
| TGEX™-HC-hG1-Zeo   | MX026   | Human IgG1           | Heavy chain expression plasmid for human IgG1   |
| TGEX™-FH-hG1-Zeo   | MX023   | Human IgG1 CH1       | Expression plasmid for human IgG1 Fab fragments   |
| TGEX™-FC-hG1-Zeo   | MX025   | Human IgG1 Fc        | Expression plasmid for human IgG1 Fc fusions  |
| TGEX™-HC-hG2-Zeo   | MX027   | Human IgG2           | Heavy chain expression plasmid for human IgG2   |
| TGEX™-HC-hG3-Zeo   | MX028   | Human IgG3           | Heavy chain expression plasmid for human IgG3   |
| TGEX™-HC-hG4[S228P]-Zeo                                      | MX029   | Human IgG4[S228P]    | Heavy chain expression plasmid for human IgG4   |
| TGEX™-HC-mG1-Zeo   | MX032   | Murine IgG1          | Heavy chain expression plasmid for murine IgG1  |
| TGEX™-HC-mG2a-Zeo  | MX033   | Murine IgG2a         | Heavy chain expression plasmid for murine IgG2a   |
| TGEX™-HC-mG2b-Zeo  | MX034   | Murine IgG2b         | Heavy chain expression plasmid for murine IgG2b   |
| TGEX™-HC-mG3-Zeo   | MX035   | Murine IgG3          | Heavy chain expression plasmid for murine IgG3  |
| TGEX™-HC-rbG-Zeo   | MX039   | Rabbit IgG           | Heavy chain expression plasmid for rabbit IgG   |
| <b>Fc-engineered Antibody Heavy Chain Expression Vectors</b> |         |                      |   |
| TGEX™-HC-hG1[EA]-Zeo   | MX041   | Human IgG1           | IgG1 heavy chain with mutated Fc to increase ADCC/CDC                                   |
| TGEX™-HC-hG1[NA]-Zeo   | MX042   | Human IgG1           | IgG1 heavy chain with mutated Fc for aglycosylated antibody                             |
| TGEX™-HC-hG1[LALA-PG]-Zeo                                    | MX043   | Human IgG1           | IgG1 heavy chain with mutated Fc to reduce ADCC/CDC                                     |
| TGEX™-HC-hG1[YTE-KF]-Zeo                                     | MX044   | Human IgG1           | IgG1 heavy chain with mutated Fc to increase serum half-life                            |
| TGEX™-HC-hG4[SPLE-PG]-Zeo                                    | MX045   | Human IgG4           | IgG4 heavy chain with mutated Fc to reduce ADCC/CDC                                     |
| <b>Antibody Light Chain Expression Vectors</b>               |         |                      |   |
| TGEX™-LC-hK-Zeo  | MX030   | Human Kappa          | Light chain expression plasmid for human Kappa  |
| TGEX™-LC-hL-Zeo  | MX031   | Human Lambda 2       | Light chain expression plasmid for human Lambda 2                                       |
| TGEX™-LC-mK-Zeo  | MX036   | Murine Kappa         | Light chain expression plasmid for murine Kappa   |
| TGEX™-LC-mL1-Zeo   | MX037   | Murine Lambda 1      | Light chain expression plasmid for murine Lambda 1                                      |
| TGEX™-LC-mL2-Zeo   | MX038   | Murine Lambda 2      | Light chain expression plasmid for murine Lambda 2                                      |
| TGEX™-LC-rbKb4   | MX040   | Rabbit Kappa b4      | Light chain expression plasmid for rabbit Kappa b4                                      |
| <b>Engineering Vectors</b>                                   |         |                      |   |
| TGEX™-AC- Zeo  | MX020   | Any expressions      | Universal mammalian expression vector   |
| TGEX™-eGFP- Zeo  | MX022   | Transfection control | Control plasmid for monitoring transient transfections                                  |
| TGEX™-SCblue-Zeo   | MX024   | scFv cloning vector  | For the transfer of scFv from any pADL phagemid vector and expression as scFv-Fc fusion |

## Examples for Desired Antibody Format

| COMBINATION                        | FORMAT                                | PURIFICATION METHOD <sup>1</sup> |
|------------------------------------|---------------------------------------|----------------------------------|
| TGEX™-HC-hG1-Zeo + TGEX™-LC-hK-Zeo | Full-length human or chimeric IgG1/K  | Protein A or G                   |
| TGEX™-FH-hG1-Zeo + TGEX™-LC-hK-Zeo | Human or chimeric IgG1/K Fab fragment | Protein L, G or IMAC             |
| TGEX™-FC-hG1                       | Human IgG1 Fc fusion                  | Protein A or G                   |
| TGEX™-SCblue-Zeo                   | Human IgG1 scFv-Fc fusion             | Protein A or G                   |

<sup>1</sup>Purification by Protein A, G, and L may require additional testing.

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